COMMUNITY ECOLOGY - ORIGINAL RESEARCH



The impact of plant chemical diversity on plant-herbivore interactions at the community level

Diego Salazar^{1,2} · Alejandra Jaramillo³ · Robert J. Marquis²

Received: 2 December 2015 / Accepted: 4 April 2016 / Published online: 29 April 2016 © Springer-Verlag Berlin Heidelberg 2016

Abstract Understanding the role of diversity in ecosystem processes and species interactions is a central goal of ecology. For plant–herbivore interactions, it has been hypothesized that when plant species diversity is reduced, loss of plant biomass to herbivores increases. Although long-standing, this hypothesis has received mixed support. Increasing plant chemical diversity with increasing plant taxonomic diversity is likely to be important for plant–herbivore interactions at the community level, but the role of chemical diversity is unexplored. Here we assess the effect of volatile chemical diversity on patterns of herbivore damage in naturally occurring patches of *Piper* (Piperaceae) shrubs in a Costa Rican lowland wet forest. Volatile chemical diversity negatively affected total, specialist, and generalist herbivore damage. Furthermore, there were

Communicated by Richard Karban.

Electronic supplementary material The online version of this article (doi:10.1007/s00442-016-3629-y) contains supplementary material, which is available to authorized users.

 Diego Salazar diegosalazar@berkeley.edu
Alejandra Jaramillo

> Robert J. Marquis Robert_Marquis@umsl.edu

alejandra.jaramillo@gmail.com

- ¹ Department of Integrative Biology, University of California-Berkeley, 1005 Valley Life Sciences Building #3140, Berkeley, CA 94720-3140, USA
- ² Department of Biology and the Whitney R. Harris World Ecology Center, One University Boulevard, University of Missouri-St. Louis, St. Louis, MO 63121, USA
- ³ Facultad de Ciencias Basicas, Universidad Militar Nueva Granada, Bogota, Colombia

differences between the effects of high-volatility and lowvolatility chemical diversity on herbivore damage. Highvolatility diversity reduced specialist herbivory, while lowvolatility diversity reduced generalist herbivory. Our data suggest that, although increased plant diversity is expected to reduce average herbivore damage, this pattern is likely mediated by the diversity of defensive compounds and general classes of anti-herbivore traits, as well as the degree of specialization of the herbivores attacking those plants.

Keywords Plant interactions · Herbivory · *Piper* · Volatile compounds · La Selva

Introduction

Community ecologists have struggled to find rules governing both the assembly of communities they study and the processes that occur within those communities (Lawton 1999; Simberloff 2004; McGill et al. 2006; Ricklefs 2008). For plant communities specifically, the relationship between plant diversity, herbivore diversity and abundance, and herbivory has been studied extensively in agricultural systems and natural habitats (Andow 1991). These studies have shown that when plant species richness is reduced, loss of primary production to herbivores generally increases (Brown and Ewel 1987; Andow 1991; Hillebrand and Cardinale 2004; Hooper et al. 2005; Jactel and Brockerhoff 2007). This pattern is most commonly explained by the "resource concentration hypothesis" (Root 1973), which maintains that a particular resource will have a lower relative abundance in diverse plant assemblages than in less diverse communities. In general, as the diversity of host plants in a community decreases, the encounter rate between a specialist herbivore and its plant host will increase, resulting in higher herbivory. Nevertheless, many studies have found that herbivore damage can also increase with higher taxonomical diversity (e.g. Scherber et al. 2006; Loranger et al. 2014). For example, Schuldt et al. (2010) compared insect herbivory on tree sapling and shrub species within 27 forest plots in southeast China, and showed that insect herbivory increased with increasing plant species richness, even after accounting for potentially confounding variables—exactly opposite that predicted by the resource concentration hypothesis.

One possible explanation for such contradictory findings is the use of species diversity as a proxy for habitat complexity. Although changes in taxonomic plant diversity are expected to affect plant-herbivore interactions, this effect is likely mediated by changes in the diversity of specific plant traits that affect the relationship between herbivores and plant hosts. Therefore, to assess the effect of diversity on plant-herbivore interactions, it is important to measure the diversity of biologically meaningful traits in addition to species diversity.

One such set of traits is plant secondary chemistry. There are two possible mechanisms by which plant chemical diversity could affect plant herbivore interactions. First, given that all herbivore species are probably limited in the number of defense chemical compounds they can overcome or circumvent, communities with high chemical diversity will likely have fewer hosts that are chemically compatible with specific herbivores. Second, in the more chemically diverse communities, the search patterns of insect herbivores could be affected by the higher semiochemical complexity. For example, for herbivores searching for host plants via volatile compounds, increases in local chemical diversity, rather than species diversity per se, could reduce the encounter rate between herbivores and host plants due to chemical disorientation and odor masking (e.g. the semiochemical redundancy hypothesis, Randlkofer et al. 2010; Beyaert and Hilker 2014).

The effect of plant diversity on plant-herbivore interactions may also depend on local plant phylogenetic diversity. Phylogenetic diversity can be defined as a measure of community complexity that incorporates the phylogenetic relationships between local species. A community with low phylogenetic diversity will comprise closely related species, and a community with high phylogenetic diversity will primarily have distantly related taxa. Therefore, this measure of diversity can be important for understanding the relationship between diversity and plant-herbivore interactions, as it can determine the similarity of phylogenetically conserved traits within the plant community. Consequently, phylogenetic diversity can also serve to capture some of the local variation of plant traits that are otherwise not explicitly measured. For phytochemical diversity specifically, it is generally expected that the closer the relationship between plant species, the more similar their chemistry will be. However, within a plant genus, herbivore selection may result in chemical diversification even among sister species, such that their differences are greater than those among more distantly related congeners (Coley and Kursar 2014).

Herbivore diet breadth is another commonly overlooked but extremely important component (Andow 1991; Specht et al. 2008; Schuldt et al. 2010) in assessing the relationship between plant diversity and plant-herbivore interactions. On one hand, herbivores with a narrow diet breadth (specialists) will have higher success finding hosts in simple, low-diversity habitats where compatible plant species are concentrated. On the other hand, herbivores with a wide diet breadth (generalists) are less likely to be affected by changes in the abundance of particular plant species, and therefore will be less sensitive to changes in local plant diversity (Root 1973; Andow 1991). Within the context of chemical diversity, herbivores with a narrow chemical diet breadth will be less likely to encounter chemically compatible host plants as the chemical complexity of a community increases.

Furthermore, it is important to note that not all secondary plant metabolites act similarly on all insect herbivores (Lankau 2007; Orians and Ward 2010; Ali and Agrawal 2012). For example, specialist herbivores are better adapted to circumvent, metabolize, or sequester specialized lowvolatility plant defenses (e.g. alkaloids, cyanides, and cardenolides); therefore, changes in the diversity of such compounds are more likely to affect generalist herbivores (Barker et al. 2002; Macel and Vrieling 2003; Macel et al. 2005; Lankau 2007; Petschenka and Agrawal 2016). Similarly, due to their narrow diet range, specialist herbivores are more likely to search for specific plant species and thus depend strongly on long-range chemical cues to locate their hosts (e.g. mono and sesquiterpenes, and phenylpropanoids; De Bruyne and Baker 2008; Bruce and Pickett 2011; Reudler et al. 2011; Büchel et al. 2014). Consequently, it is reasonable to expect that changes in the diversity of highvolatility compounds will have a stronger effect on specialist than on generalist herbivores.

In this study, we assessed the effect of plant secondary chemistry on plant-herbivore interactions at the community level, taking into account herbivore diet breadth and plant phylogenetic relationships, for the neotropical species-rich genus *Piper* (Piperaceae). We measured the effect of chemical diversity on total, specialist, and generalist herbivore damage on a set of naturally occurring *Piper* communities in a lowland tropical wet forest. In addition, we assessed how the diversity of different secondary compounds (highvolatility vs. low-volatility) affected levels of specialist and generalist herbivory. We predicted that secondary compounds of high-volatility would have a greater impact on specialist herbivore species than on generalists, because the former are more reliant on volatiles to find their host plants. In contrast, generalists would be more strongly affected by low-volatility compounds than would specialists, which are likely to be better adapted to cope with such compounds.

Materials and methods

Study site and system

The study was conducted at the La Selva Biological Station in Costa Rica (owned and operated by the Organization for Tropical Studies) between May and August 2007. The station is located in the Atlantic lowlands of Puerto Viejo de Sarapiquí (Heredia). It contains more than 1600 ha of tropical wet forest and receives approximately 4000 mm of rainfall annually. To date, approximately 1850 species-rich of plants have been found in La Selva, 50 of which are in the genus *Piper*.

Piper (Piperaceae) is a pantropical genus with approximately 1000 species in the neotropics (Jaramillo 2006). The natural range of the genus in the New World extends from northern Mexico to northern Argentina. Piper species are abundant in low- and mid-elevation forests, rarely reaching 2500 m, and are often among the most species-rich plant genera in neotropical forests (Gentry 1982; Marquis 2004). Most Piper species at La Selva occur in discrete patches that can contain up to 21 different species (Salazar et al. 2013). Piper secondary metabolite diversity has been studied extensively, and there is an important body of published methods for compound isolation, chemical profiling, and artificial synthesis. Furthermore, the effects of Piper compounds on herbivores, pathogens, and seed dispersal are well documented (Parmar et al. 1997; Dyer et al. 2001; Mikich et al. 2003; Dyer et al. 2004; Kato and Furlan 2007; Fincher et al. 2008; Marques et al. 2010).

Piper communities and herbivore damage

To assess the effect of *Piper* chemical diversity on herbivore damage, we sampled 81 naturally occurring multispecies patches of *Piper* throughout the primary forest of La Selva. Patches were selected by performing transects parallel to the station trails (transects were between 50 and 100 m from the trail). All patches selected were at least 250 m apart. To standardize sampling, a plot with a 10 m radius was set up in every patch. In each plot we counted the number of *Piper* individuals with a stem diameter of 1 cm or greater at ground level; all *Piper* plants were identified to species.

For each *Piper* present in a plot, we visually assessed the percentage of specialist and generalist herbivory, measured as the percentage of leaf area removed. Every plant was assigned a value between 0 and 95 %, in increments of 5 %, for each herbivory type. Specialist and generalist herbivory in all *Piper* plants were easily distinguishable by the characteristic skeletonization of leaves by *Piper* specialist herbivores of the genus *Eois* (Geometridae) (Connahs et al. 2009; Dyer et al. 2010). The neotropical species of this genus are highly specialized and feed on one or rarely two species of *Piper* (Connahs et al. 2009). There are other *Piper* specialists at La Selva (e.g. weevils and leaf beetles; Marquis 1990); however, damage by these species was rarely encountered. Furthermore, *Eois* caterpillars have accounted for up to 75 % of the damage observed in *Piper* species (Salazar et al. 2013). For this reason, all non-*Eois* damage was assigned to generalist herbivores.

To assess the effect of chemical diversity independently of taxonomic species diversity, we calculated each plot's taxonomic diversity (hereafter, Piper diversity) using the Gini-Simpson index. To account for the effect that the surrounding plant diversity could have on Piper herbivore damage, we also counted and identified all non-Piper plants present within all plots (hereafter, non-Piper diversity). Non-Piper diversity was calculated for each plot using the Gini-Simpson index as well. Diversity indices were calculated using EcoSim 7.1 software (Gotelli and Entsminger 2012). Finally, given that light availability can influence leaf quality and palatability, and thus may affect levels of leaf herbivory (Angulo-Sandoval and Aide 2000; Blundell and Peart 2001; Takafumi et al. 2010; Salgado-Luarte and Gianoli 2011), we measured canopy openness (hereafter, light) for each plot using hemispherical photography. This technique measures the percentage of canopy cover using fisheye photographs of the forest canopy and the Gap Light Analyzer (GLA) 2.0 software package (Frazer et al. 1999).

Chemical analysis

For all Piper species, we collected leaf material samples from young, fully expanded leaves with 5-10 % herbivore damage (to control for the effect of induction). At least four samples were collected for each species, all from different plant individuals and different transects (plants were randomly selected within each transect). Samples were dried with silica gel and transported to the University of Missouri-St. Louis for chemical analysis. From each sample, 0.4 g of dried material was ground to a fine powder under liquid nitrogen. To extract a broad range of secondary metabolites (polar and non-polar), samples were extracted using 1.5 ml of 1:1 methanol-chloroform solution. Samples were kept at temperatures below 5 °C at all times to reduce the loss of volatile compounds. As an internal standard, 0.1 mg of piperine was added to all samples. Samples were filtered (0.2μ) and stored in volatile organic chemical (VOC) vials at -80 °C until analysis. Qualitative chemical analysis of the extracts was performed using GC-MS (HP 5890 coupled with a quadrupole model 5988A mass detector) with helium as a carrier gas and an HP-5 capillarv column (30, 0.32 mm ID, 0.25 µm). To ensure good chromatographic resolution and to reduce the likelihood of co-eluting compounds, we used very slow runs (80 min). It is important to note that although this study focuses only on compounds that are detectable with GC-MS, an analysis of more than 3500 records of secondary compounds found in Piper from NAPRALERT (Natural Products Alert Database; Loub et al. 1985) shows that these compounds account for more than 75 % of all Piper secondary metabolites reported in this database (Fig. S1). Because the abundance of the secondary compounds can vary among individuals due to factors such as induction, genetic variability, and resource availability, we used only presence and absence data of chromatographic features. To assess chemical dissimilarity among all sample species, we used mass spectra and retention-time chromatogram alignment. Chromatograms were aligned using MZmine (Pluskal et al. 2010). The mass spectra of the different compounds in the samples were compared with NIST/EPA/NIH and MassBank (Horai et al. 2010) databases and with the primary literature. Metabolites that did not match the available sources and databases were classified as unknown. It is also important to note that this analytical approach does not depend on the precise identification of chemical compounds. Here, a combination of mass spectral patterns, molecular mass, and retention times was used to ascertain the commonality of chemical compounds across plant species (see the proof of concept section in the Supporting Information).

Plot chemical diversity

To assess the chemical diversity of the 81 sampled plots, we first used the pvclust function from the pvclust R package (Suzuki and Shimodaira 2006; R Core Team 2012) to generate chemical dendrograms (hierarchical clustering analyses; Ward's algorithm) based on chemical data. We generated three different dendrograms: (1) a species total chemical dendrogram, (2) a species high-volatility chemical dendrogram, including only low molecular weight (<230 g/mol) terpenoids and phenylpropanoids, and (3) a species low-volatility but high molecular weight (>250 g/ mol) chemical dendrogram (amides, flavonoids, lignans, catechols, and cinnamic acids). We subsequently extracted the species pair matrices of chemical distances from the dendrograms (cophenetic function from stats package). Finally, we used the chemical distance matrices of these three species to calculate the total, high-volatility, and low-volatility chemical diversity for all sampled plots. All values were assessed using Rao's quadratic entropy index with the raoD function in the Picante package for R (Rao 2010; Ricotta and Moretti 2011; Kembel et al. 2012). We used Rao's index for two main reasons. First, Rao takes into account the species chemical differences: a community with a high Rao chemical diversity will comprise plants that are very different in chemical composition, and a community with low Rao chemical diversity will have species with similar chemical composition. Second, Rao can also account for the differences in relative abundance of *Piper* species within each plot.

Phylogenetic diversity

To control for the potential effect of phylogeny on Piper herbivory and chemical similarity, we constructed a phylogenetic tree of all sampled Piper species. Samples of leaves were collected to perform sequencing of the ITS and the chloroplast intron psbJ-petAx for phylogenetic analysis (following Jaramillo et al. 2008). The resulting phylogeny was concurrent with all current phylogenetic and taxonomic data (Fig. S2). We used this phylogenetic reconstruction to calculate the Rao phylogenetic diversity index of each plot, again using the Picante package. Similar to the chemical indices, Rao's phylogenetic diversity takes into account species phylogenetic distances and species relative abundance. Our measures of plant (Gini-Simpson) and phylogenetic and chemical diversity (Rao quadratic index) are ones that are relevant to the searching herbivore, i.e. they measure the probability that the searching herbivore will encounter a new plant species based on chemical and phylogenetic similarity.

Statistical analysis

We analyzed the data using generalized linear mixed effects models (GLMM). To test the role of chemical diversity on Piper herbivory, we assessed the effect of total chemical diversity, Piper phylogenetic diversity, and non-Piper diversity (as fixed effects) of each *Piper* patch on total, generalist, and specialist herbivory suffered by each Piper plant (models A.1, A.2, and A.3, respectively; see Table S1 in Supporting Information). In all of these models, we controlled for light and Piper diversity (random effects). We also assessed the effect of high-volatility diversity, lowvolatility diversity, Piper phylogenetic diversity, and non-Piper diversity (as fixed effects) on Piper generalist and specialist herbivory (models B.1 and B.3, respectively), again controlling for light and Piper diversity (random effects). In all models, the experimental unit was the Piper plot. We also evaluated three models to assess the effect of Piper taxonomic diversity on herbivore damage (Table S2, models C.1, C.2, and C.3). Model fit was evaluated via the Akaike information criterion (AIC) and maximum

likelihood tests (*see* Supporting Information for details on model construction and model selection). All models used the maximum likelihood estimation method, and models

Table 1 Results from the generalized linear mixed model for the optimal models

	Estimate	df	t	p value
Model A.1				
Total herbivory = total chemical diversity)	diversity +	(light) + (<i>Pip</i>	er
Total chemical diversity*	-0.06	1, 79	-3.94	0.0001
Model A.2				
Generalist herbivory = total chemical diversity + <i>Piper</i> phyloge- netic diversity + non- <i>Piper</i> diversity + (light) + (<i>Piper</i> diversity)				
Total chemical diversity*	-0.02	1,27	-3.84	0.0006
Piper phylogenetic diversity	0.005	1, 72	1.4	0.16
Non-Piper diversity	-0.51	1, 76	-1.04	0.30
Model A.3				
Specialist herbivory = total chem diversity)	ical divers	ity + (light) +	(Piper
Total chemical diversity*	-0.07	1, 79	-4.54	< 0.0001
Model B.1				
Specialist herbivory = high-volatility chemical diversity + <i>Piper</i> phylogenetic diversity + (light) + (<i>Piper</i> diversity)				
High-volatility chemical diver- sity*	-0.14	1, 78	-2.34	0.02
Piper phylogenetic diversity*	-0.09	1,78	-2.12	0.03
Model B.2				
Generalist herbivory = low-volatility chemical diversity + (light) + (<i>Piper</i> diversity)				
Low-volatility chemical diver- sity*	-0.18	1, 75	-2.91	0.004

Complete models and details of model selection procedure can be found in the supplementary information. Random variables are shown in parentheses. Variables with an *asterisk* (*) show a statistically significant effect (p < 0.05)

were analyzed using R 2.15 (R Core Team 2012) and the nlme package with the functions lme and anova (Pinheiro et al. 2016). Finally, given that different measures of diversity can be correlated, all beyond-optimal models were checked for the possible effect of multicollinearity on our analysis by calculating the variable inflation factor (VIF).

Results

After controlling for light and species diversity, the final optimal generalized linear mixed model showed that the total chemical diversity in a plot had a significant negative effect on total herbivore damage; higher levels of chemical diversity within a plot were associated with lower levels of herbivore damage (model A.1; VIF < 2.4; Tables 1, S1; Fig. 1a). The mean leaf area loss in the most chemically diverse and least chemically diverse plots was 4.5 and 11.1 %, respectively. An identical pattern was found for the effect of total chemical diversity on specialist herbivory (model A.3; VIF < 2.2, Fig. 1c). The final model for generalist herbivory included non-Piper diversity, Piper phylogenetic diversity, and Piper total chemical diversity (model A.2; VIF < 2.4; see Tables 1, S1). Only total chemical diversity, however, had a significant effect on generalist herbivory (model A.2; Table 1; Fig. 1b).

High-volatility chemical diversity had a significant negative effect on specialist herbivory (model B.1; VIF < 1.2; Tables 1, S1; Fig. 2). In contrast, higher levels of low-volatility diversity had a significant negative effect on generalist herbivory (model B.2; VIF < 1.2; Tables 1, S1; Fig. 2). The model selection procedure showed that high-volatility chemical diversity did not improve the model explaining generalist herbivory (model B.2), and that low-volatility chemical diversity did not improve the model explaining specialist herbivore damage (model B.1, Tables 1, S1; Fig. 2). Furthermore, adding phylogenetic *Piper* diversity



Fig. 1 The different panes show the relationship between total (**a**), generalist (**b**), and specialist (**c**) herbivore damage (percentage of leaf area removed) and total *Piper* volatile chemical diversity (Rao's

index). Each point represents an experimental plot. *Grey lines* are linear regressions, and the values for the regressions are shown in the *upper-right corner* of each *plot*

Fig. 2 The different panes show the relationship between high- and low-volatility chemical diversity (Rao's index) and generalist and specialist herbivore damage (percentage of leaf area removed). Each point represents an experimental plot. Grey lines are linear regressions, and the values for the regressions are shown in the upper-*right* corner of each plot. a Percentage of generalist herbivory vs. high-volatility chemical diversity. b Percentage of generalist herbivory versus low-volatility chemical diversity. c Percentage of specialist herbivory vs. high-volatility chemical diversity. d Percentage of specialist herbivory vs. lowvolatility chemical diversity



improved only the model that explained specialist herbivory (model B.2; Tables 1, S1). *See* the general results from the plant, herbivory, and chemical surveys in the Supporting Information.

Discussion

The overall results from this study suggest that communitywide levels of chemical diversity influence plant-herbivore interactions. We found that plots with higher levels of total *Piper* chemical diversity were associated with lower levels of total, generalist, and specialist insect herbivory. Furthermore, this pattern held true after controlling for non-Piper diversity, Piper taxonomic diversity, light availability, and *Piper* phylogenetic relationships. In addition, we found that different kinds of chemical diversity affected specific guilds of insect herbivores differentially, in this case specialist and generalist herbivores. Plots with higher levels of high-volatility chemical diversity had significantly lower specialist insect herbivory, while plots with higher low-volatility chemical diversity showed lower levels of damage caused by generalist insect herbivores. Thus, the Piper community composition growing around an individual Piper plant at the study site influenced the amount of damage that the plant received from its herbivores. This neighborhood effect is apparently influenced mainly by the presence of conspecifics and congeners, not by plants from other families.

We show here that the effects of neighborhood chemical diversity on the amount of herbivore damage appear to be independent of taxonomic and phylogenetic plant diversity. Two mechanisms that might explain the effects of chemical diversity on the local plant herbivore damage found in this study are (1) the resource concentration principle (Root 1973) and (2) the semiochemical-diversity hypothesis (Zhang and Schlyter 2003; Randlkofer et al. 2010).

Resource concentration hypothesis

Root's resource concentration hypothesis states that the amount of herbivore damage experienced by a given plant will depend on the encounter rate between the plant and its herbivores. In more diverse plant assemblages, a particular plant host will have a lower relative abundance than in a simpler, less diverse community. Insect herbivores have been reported to feed upon tissue containing a limited set of plant secondary compounds, due to evolutionary (Becerra 1997; Becerra and Venable 1999), metabolic (Freeland and Janzen 1974; Scriber 2002; Marsh et al. 2006), and behavioral constraints (Fernandez and Hilker 2007; Schröder and Hilker 2008; Cheng et al. 2013). Therefore, it is also likely that an increase in local plant chemical diversity will reduce the encounter rate between the insect herbivore and chemically compatible plant hosts, resulting in a comparable reduction in the plant herbivore damage.

Another prediction that can be made from Root's hypothesis is that changes in chemical diversity will have a stronger effect on herbivores that feed on a small set of plant secondary metabolites (specialists) than on generalists who are able to feed on many plant species with very diverse defense chemistry. Thus, the significant effect of total plant chemical diversity on specialist herbivores found in this study supports this hypothesis.

Semiochemical-diversity hypothesis

Herbivore chemical disorientation due to local plant chemical complexity can also influence plant-herbivore interactions. The semiochemical-diversity hypothesis (Zhang and Schlyter 2003) states that complex plant communities will have greater secondary compound diversity than simpler communities, and that these higher levels of chemical diversity are likely to pose a challenge to herbivore orientation, by negatively affecting their ability to find, recognize, or accept a particular plant host (for similar ideas see Schröder and Hilker 2008; Dicke and Baldwin 2010; Jactel et al. 2011; Party et al. 2013; Beyaert and Hilker 2014). For example, combinations of specific plant odors can mask host chemical cues or even repel insect herbivores (Party et al. 2013). By interfering with the herbivore's ability to track critical plant-host chemical cues, higher levels of plant chemical diversity could strongly affect herbivorehost encounter rates, thus reducing the local levels of plant damage caused by herbivores. Our results are consistent with this hypothesis.

Another prediction that can be drawn from this hypothesis is that the chemical disorientation effect of higher chemical diversity is likely to have a stronger effect on herbivores that rely heavily on volatile secondary metabolites for finding suitable hosts. In the case of *Piper* herbivores, we do not know which species use volatiles to find their host plants, but our results suggest that at least a subset of *Piper* herbivores do so (specialists). It is also important to note that these two mechanisms (resource dilution and chemical disorientation) are not mutually exclusive, and that both could act simultaneously in natural systems.

Chemical diversity and herbivore natural enemies

Another mechanism that has been proposed to explain the relationship between diversity and plant–herbivore interactions is the enemy hypothesis (Root 1973). This hypothesis states that habitats with higher plant diversity can sustain or attract a higher diversity of herbivore predators and parasitoids. Although it is difficult to imagine how chemical diversity *per se* could directly affect the diversity of generalist and opportunistic herbivore consumers, the chemical disorientation effect could alter the ability of

the specialized natural enemies of herbivores (particularly hymenopteran parasitoids) to find their prey (Wäschke et al. 2013). For *Piper* herbivores specifically, *Eois* caterpillars, for example, are frequently parasitized, resulting in significant mortality rates (Connahs et al. 2009; Richards et al. 2010). Nevertheless, recent studies have suggested that parasitoids are attracted to a combination of plant and herbivore volatile metabolites, and are therefore less affected by changes in plant species and chemical diversity alone (Wäschke et al. 2014; Kruidhof et al. 2015). In order to determine the net effect of chemical disorientation on plant herbivore damage—directly on herbivore attack and indirectly on parasitism—more specifically designed laboratory and field experiments are needed.

Effects of low- and high-volatility chemical diversity

After partitioning the plot's chemical diversity into highvolatility and low-volatility groups, it was possible to assess the effect of specific groups of secondary metabolites on plant herbivore damage. These two groups of secondary compounds showed markedly contrasting effects on generalist and specialist herbivore damage.

Specialist herbivores are generally better adapted to circumventing or overcoming low-volatility secondary plant metabolites (e.g. alkaloids and glucosinolates; Orians 2000; Lankau 2007). Therefore, it is reasonable to expect that these herbivores will not be especially susceptible to changes in the diversity of low-volatility compounds. Furthermore, specialist herbivores are expected to feed on a small subset of Piper species within our plots (Connahs et al. 2009; Dyer et al. 2010), and it is thus unlikely that the presence of different low-volatility compounds in the surrounding non-host plants could affect the amount of damage these specialist insects cause (Macel et al. 2002; Macel and Vrieling 2003; Cheng et al. 2013). In contrast, despite the fact that specialist herbivores are also expected to have highly developed host search mechanisms, it is likely that higher levels of community-wide high-volatility chemical diversity can affect their ability to track the specific chemical cues from their hosts, due to odor masking and semiochemical redundancy (Schröder and Hilker 2008; Randlkofer et al. 2010; Bruce and Pickett 2011; Party et al. 2013; Zakir et al. 2013).

In contrast, generalist herbivores feed on a wide range of plants, and are less likely to track specific high-volatility chemical cues from a particular host plant in order to find a suitable host (e.g. Vargas et al. 2005). Thus, changes in the community's high-volatility diversity are less likely to affect their ability to find a suitable plant for feeding. Instead, low-volatility diversity was found to negatively affect generalist herbivore damage. Researchers have reported that generalist herbivores are less able to cope with specialized defenses of a particular plant host (Van Dam et al. 1995; Macel 2011; Ali and Agrawal 2012; Lampert 2012), and this is true for some *Piper* generalist herbivores (Richards et al. 2010). Given that most of the low-volatility secondary compounds found in this study included alkaloids (including amides and imides), flavo-noids, and catechols, it is reasonable to expect that a higher diversity of such compounds could reduce the number of palatable hosts available to generalist herbivores.

The mechanisms linking chemical diversity to either generalist or specialist herbivory appear to differ. On the one hand, our data suggest that the effect of chemical diversity on the reduction of specialist herbivore damage is the result of semiochemical saturation, redundancy, or odor masking (semiochemical diversity hypothesis). On the other, generalist herbivores appear to be affected by lowvolatility diversity. Given that low-volatility compounds are less likely to be used as cues for host-finding by generalist herbivores, this pattern suggests that the effect of high lowvolatility diversity on generalist herbivory is the result of "resource dilution" (Root's hypothesis). Furthermore, the different effects of high-volatility and low-volatility chemistry on different guilds of herbivores could also explain the strong difference in the patterns of high-volatility and lowvolatility similarity across Piper species (Fig. S3).

Finally, it is important to note that the inclusion of Piper phylogenetic diversity did not improve most of the models used for this study. This result is likely because Piper chemical diversity does not show a significant phylogenetic signal for the 27 Piper species found in our plots (Salazar et al. 2013). It is generally assumed that insect herbivores are more likely to feed on closely related plant species, because these species are more likely to share similar traits than are distantly related species. When this assumption does not hold true (see also Kursar et al. 2009), community phylogenetic complexity will not necessarily have a significant effect on the encounter rate between herbivore and chemically compatible plant hosts. Lastly, the small effect that phylogenetic diversity had on specialist herbivory could either be (1) linked to phylogenetically conserved anti-herbivore traits not measured in this study or (2) evidence of a coevolutionary history between Piper and their specialized herbivores (Eois).

Conclusions

Our results suggest that interactions between *Piper* and its natural enemies at the community level are significantly affected by local secondary metabolite diversity. These results also underscore the importance of studying different biologically relevant dimensions of community complexity when assessing the relationship between diversity and plant-herbivore interactions. Different measures of diversity will likely be associated with different herbivore guilds and taxonomic groups. Additionally, our results also suggest that these patterns can be mediated by multiple mechanisms acting in concert (e.g. resource concentration and semiochemical-diversity). Whether these results apply to other diverse tropical genera, or even temperate genera such as *Solidago*, *Salix*, and *Quercus*, remains to be seen.

Acknowledgments We thank the staff of the Organization for Tropical Studies for their logistical support and two anonymous reviewers for comments on an earlier draft. Financial support came from the Whitney R. Harris World Ecology Center, Elizabeth Kellogg Lab, the Organization for Tropical Studies, and the National Science Foundation (DEB-1210643).

Author contribution statement DS and RJM conceived and designed the study. DS collected the data and performed the chemical analysis. AJ performed phylogenetic analysis. DS, RJM and AJ wrote manuscript.

References

- Ali JG, Agrawal AA (2012) Specialist versus generalist insect herbivores and plant defense. Trends Plant Sci 17:293–302
- Andow DA (1991) Vegetational diversity and arthropod population response. Annu Rev Entomol 36:561–586
- Angulo-Sandoval P, Aide TM (2000) Effect of plant density and light availability on leaf damage in *Manilkara bidentata* (Sapotaceae). J Trop Ecol 16:447–464
- Barker A, Schaffner U, Boevé JL (2002) Host specificity and host recognition in a chemically-defended herbivore, the tenthredinid sawfly *Rhadinoceraea nodicornis*. Entomol Exp Appl 104:61–68
- Becerra JX (1997) Insects on plants: macroevolutionary chemical trends in host use. Science 276:253–256
- Becerra JX, Venable DL (1999) Macroevolution of insect-plant associations: the relevance of host biogeography to host affiliation. Proc Natl Acad Sci USA 96:12626–12631
- Beyaert I, Hilker M (2014) Plant odour plumes as mediators of plantinsect interactions. Biol Rev 89:68–81
- Blundell AG, Peart DR (2001) Growth strategies of a shade-tolerant tropical tree: the interactive effects of canopy gaps and simulated herbivory. J Ecol 89:608–615
- Brown BJ, Ewel JJ (1987) Herbivory in complex and simple tropical successional ecosystems. Ecology 68:108–116
- Bruce TJA, Pickett JA (2011) Perception of plant volatile blends by herbivorous insects—finding the right mix. Phytochemistry 72:1605–1611
- Büchel K, Austel N, Mayer M, Gershenzon J, Fenning TM, Meiners T (2014) Smelling the tree and the forest: elm background odours affect egg parasitoid orientation to herbivore induced terpenoids. Biocontrol 59:29–43
- Cheng D, van der Meijden E, Mulder PPJ, Vrieling K, Klinkhamer PGL (2013) Pyrrolizidine alkaloid composition influences cinnabar moth oviposition preferences in *Jacobaea* hybrids. J Chem Ecol 39:430–437
- Coley PD, Kursar TA (2014) On tropical forests and their pests. Science 343:35–36
- Connahs H, Rodríguez-Castañeda G, Walters T, Walla T, Dyer LA (2009) Geographic variation in host-specificity and parasitoid

pressure of an herbivore (Geometridae) associated with the tropical genus *Piper* (Piperaceae). J Insect Sci 9:28

- De Bruyne M, Baker TC (2008) Odor detection in insects: volatile codes. J Chem Ecol 34:882–897
- Dicke M, Baldwin IT (2010) The evolutionary context for herbivoreinduced plant volatiles: beyond the 'cry for help'. Trends Plant Sci 15:167–175
- Dyer LA, Dodson CD, Beihoffer J, Letourneau DK (2001) Trade-offs in antiherbivore defenses in *Piper cenocladum*: ant mutualists versus plant secondary metabolites. J Chem Ecol 27:581–592
- Dyer LA, Letourneau DK, Dodson CD, Tobler MA, Stireman Iii JO, Hsu A (2004) Ecological causes and consequences of variation in defensive chemistry of a neotropical shrub. Ecology 85:2795–2803
- Dyer LA, Letourneau DK, Chavarria GV, Salazar D (2010) Herbivores on a dominant understory shrub increase local plant diversity in rain forest communities. Ecology 91:3707–3718
- Fernandez P, Hilker M (2007) Host plant location by Chrysomelidae. Basic Appl Ecol 8:97–116
- Fincher RM, Dyer LA, Dodson CD, Richards JL, Tobler MA, Searcy J, Mather JE, Reid AJ, Rolig JS, Pidcock W (2008) Inter-and intraspecific comparisons of antiherbivore defenses in three species of rainforest understory shrubs. J Chem Ecol 34:558–574
- Frazer GW, Canham CD, Lertzman KP (1999) Gap Light Analyzer (GLA), version 2.0: imaging software to extract canopy structure and gap light transmission indices from true-colour fisheye photographs, users manual and program documentation. Simon Fraser University, Burnaby, British Columbia, and the Institute of Ecosystem Studies, Millbrook, New York
- Freeland WJ, Janzen DH (1974) Strategies in herbivory by mammals: the role of plant secondary compounds. Am Nat 108:269–289
- Gentry AH (1982) Neotropical floristic diversity: phytogeographical connections between Central and South America, Pleistocene climatic fluctuations, or an accident of the Andean orogeny? Ann Missouri Bot Garden 69:557–593
- Gotelli NJ, Entsminger GL (2012) EcoSim: Null models software for ecology. Version 7.0. Acquired Intelligence Inc. & Kesey-Bear
- Hillebrand H, Cardinale BJ (2004) Consumer effects decline with prey diversity. Ecol Lett 7:192–201
- Hooper DU, Chapin Iii FS, Ewel JJ, Hector A, Inchausti P, Lavorel S, Lawton JH, Lodge DM, Loreau M, Naeem S, Schmid B, Setala H, Symstad AJ, Vandermeer J, Wardle DA (2005) Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. Ecol Monogr 75:3–35
- Horai H, Arita M, Kanaya S, Nihei Y, Ikeda T, Suwa K, Ojima Y, Tanaka K, Tanaka S, Aoshima K, Oda Y, Kakazu Y, Kusano M, Tohge T, Matsuda F, Sawada Y, Hirai MY, Nakanishi H, Ikeda K, Akimoto N, Maoka T, Takahashi H, Ara T, Sakurai N, Suzuki H, Shibata D, Neumann S, Iida T, Funatsu K, Matsuura F, Soga T, Taguchi R, Saito K, Nishioka T (2010) MassBank: a public repository for sharing mass spectral data for life sciences. J Mass Spectrom 45:703–714
- Jactel H, Brockerhoff EG (2007) Tree diversity reduces herbivory by forest insects. Ecol Lett 10:835–848
- Jactel H, Birgersson G, Andersson S, Schlyter F (2011) Non-host volatiles mediate associational resistance to the pine processionary moth. Oecologia 166:703–711
- Jaramillo MA (2006) Using *Piper* species diversity to identify conservation priorities in the Choco region of Colombia. Biodivers Conserv 15:1695–1712
- Jaramillo MA, Callejas R, Davidson C, Smith JF, Stevens AC, Tepe EJ (2008) A phylogeny of the tropical genus *piper* using ITS and the chloroplast intron psbJ-petA. Syst Bot 33:647–660
- Kato MJ, Furlan M (2007) Chemistry and evolution of the Piperaceae. Pure Appl Chem 79:529–538

- Kembel SW, Ackerly DD, Blomberg SP, Cornwell WK, Cowan PD, Helmus MR, Webb CO, Morlon H (2012) Picante, R tools for integrating phylogenies and ecology. Bioinform 29(11):1463– 1464. doi:10.1093/bioinformatics/btq166
- Kruidhof HM, Roberts AL, Magdaraog P, Muñoz D, Gols R, Vet LEM, Hoffmeister TS, Harvey JA (2015) Habitat complexity reduces parasitoid foraging efficiency, but does not prevent orientation towards learned host plant odours. Oecologia 179:353–361
- Kursar TA, Dexter KG, Lokvam J, Pennington RT, Richardson JE, Weber MG, Murakami ET, Drake C, McGregor R, Coley PD (2009) The evolution of antiherbivore defenses and their contribution to species coexistence in the tropical tree genus Inga. Proc Natl Acad Sci 106:18073–18078
- Lampert E (2012) Influences of plant traits on immune responses of specialist and generalist herbivores. Insect. 3:573–592
- Lankau RA (2007) Specialist and generalist herbivores exert opposing selection on a chemical defense. New Phytol 175:176–184
- Lawton JH (1999) Are there general laws in ecology? Oikos 84:177–192
- Loranger H, Weisser WW, Ebeling A, Eggers T, De Luca E, Loranger J, Roscher C, Meyer ST (2014) Invertebrate herbivory increases along an experimental gradient of grassland plant diversity. Oecologia 174:183–193
- Loub WD, Farnsworth NR, Soejarto DD, Quinn ML (1985) NAP-RALERT: computer handling of natural product research data. J Chem Inf Comput Sci 25:99–103
- Macel M (2011) Attract and deter: a dual role for pyrrolizidine alkaloids in plant-insect interactions. Phytochem Rev 10:75–82
- Macel M, Vrieling K (2003) Pyrrolizidine alkaloids as oviposition stimulants for the cinnabar moth, *Tyria jacobaeae*. J Chem Ecol 29:1435–1446
- Macel M, Klinkhamer PGL, Vrieling K, Van der Meijden E (2002) Diversity of pyrrolizidine alkaloids in *Senecio* species does not affect the specialist herbivore *Tyria jacobaeae*. Oecologia 133:541–550
- Macel M, Bruinsma M, Dijkstra SM, Ooijendijk T, Niemeyer HM, Klinkhamer PGL (2005) Differences in effects of pyrrolizidine alkaloids on five generalist insect herbivore species. J Chem Ecol 31:1493–1508
- Marques JV, De Oliveira A, Raggi L, Young MCM, Kato MJ (2010) Antifungal activity of natural and synthetic amides from *piper* species. J Braz Chem Soc 21:1807–1813
- Marquis RJ (1990) Genotypic variation in leaf damage in *Piper arieianum* (Piperaceae) by a multispecies assemblage of herbivores. Evolution 44:104–120
- Marquis RJ (2004) The biogeography of Neotropical *Piper*. In: Dyer L, Palmer A (eds) *Piper*: a model genus for studies of chemistry, ecology, and evolution. Kluwer Academic Press, New York, pp 199–203
- Marsh KJ, Wallis IR, Andrew RL, Foley WJ (2006) The detoxification limitation hypothesis: where did it come from and where is it going? J Chem Ecol 32:1247–1266
- McGill BJ, Maurer BA, Weiser MD (2006) Empirical evaluation of neutral theory. Ecology 87:1411–1423
- Mikich SB, Bianconi GV, Maia BHLNS, Teixeira SD (2003) Attraction of the fruit-eating bat *Carollia perspicillata* to *Piper gaudichaudianum* essential oil. J Chem Ecol 29:2379–2383
- Orians CM (2000) The effects of hybridization in plants on secondary chemistry: implications for the ecology and evolution of plantherbivore interactions. Am J Bot 87:1749–1756
- Orians CM, Ward D (2010) Evolution of plant defenses in nonindigenous environments. Ann Rev Entomol 55:439–459
- Parmar VS, Jain SC, Bisht KS, Jain R, Taneja P, Jha A, Tyagi OD, Prasad AK, Wengel J, Olsen CE, Boll PM (1997) Phytochemistry of the genus *Piper*. Phytochemistry 46:597–673

- Party V, Hanot C, Büsser DS, Rochat D, Renou M (2013) Changes in odor background affect the locomotory response to pheromone in moths. PLoS One 8(1):e52897. doi:10.1371/journal. pone.0052897
- Petschenka G, Agrawal AA (2016) How herbivores coopt plant defenses: natural selection, specialization, and sequestration. Curr Opin Insect Sci 14:17–24
- Pinheiro JC, Bates D, DebRoy S, Sarkar D, R Core Team (2016) nlme: Linear and nonlinear mixed effects models. R package version 3.1-127
- Pluskal T, Castillo S, Villar-Briones A, Orešič M (2010) MZmine 2: modular framework for processing, visualizing, and analyzing mass spectrometry-based molecular profile data. BMC Bioinform 11:395
- R Core Team (2012) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Randlkofer B, Obermaier E, Hilker M, Meiners T (2010) Vegetation complexity-the influence of plant species diversity and plant structures on plant chemical complexity and arthropods. Basic Appl Ecol 11:383–395
- Rao CR (2010) Quadratic entropy and analysis of diversity. Sankhya. Indian J Stat 72:70–80
- Reudler JH, Biere A, Harvey JA, van Nouhuys S (2011) Differential performance of a specialist and two generalist herbivores and their parasitoids on *Plantago lanceolata*. J Chem Ecol 37:765–778
- Richards LA, Dyer LA, Smilanich AM, Dodson CD (2010) Synergistic effects of amides from two *Piper* species on generalist and specialist herbivores. J Chem Ecol 36:1105–1113
- Ricklefs RE (2008) Disintegration of the ecological community. Am Nat 172:741–750
- Ricotta C, Moretti M (2011) CWM and Rao's quadratic diversity: a unified framework for functional ecology. Oecologia 167:181–188
- Root RB (1973) Organization of a plant-arthropod association in simple and diverse habitats: the fauna of collards (*Brassica oleracea*). Ecol Monogr 43:95–120
- Salazar D, Kelm DH, Marquis RJ (2013) Directed seed dispersal of *Piper* by *Carollia perspicillata* and its effect on understory plant diversity and folivory. Ecology 94:2444–2453
- Salgado-Luarte C, Gianoli E (2011) Herbivory may modify functional responses to shade in seedlings of a light-demanding tree species. Funct Ecol 25:492–499
- Scherber C, Mwangi PN, Temperton VM, Roscher C, Schumacher J, Schmid B, Weisser WW (2006) Effects of plant diversity on invertebrate herbivory in experimental grassland. Oecologia 147:489–500

- Schröder R, Hilker M (2008) The relevance of background odor in resource location by insects: a behavioral approach. Bioscience 58:308–316
- Schuldt A, Baruffol M, Bohnke M, Bruelheide H, Hardtle W, Lang AC, Nadrowski K, von Oheimb G, Voigt W, Zhou H, Assmann T (2010) Tree diversity promotes insect herbivory in subtropical forests of south-east China. J Ecol 98:917–926
- Scriber JM (2002) Evolution of insect-plant relationships: chemical constraints, coadaptation, and concordance of insect/plant traits. Entomol Exp Appl 104:217–235
- Simberloff D (2004) Community ecology: is it time to move on? (An american society of naturalists presidential address). Am Nat 163:787–799
- Specht J, Scherber C, Unsicker SB, Kohler G, Weisser WW (2008) Diversity and beyond: plant functional identity determines herbivore performance. J Anim Ecol 77:1047–1055
- Suzuki R, Shimodaira H (2006) Hierarchical clustering with P-values via multiscale bootstrap resampling. R package version 12-0
- Takafumi H, Kawase S, Nakamura M, Hiura T (2010) Herbivory in canopy gaps created by a typhoon varies by understory plant leaf phenology. Ecol Entomol 35:576–585
- Van Dam NM, Vuister LW, Bergshoeff C, De Vos H, Van Der Meijden E (1995) The raison d'etre of pyrrolizidine alkaloids in *Cynoglossum officinale*: deterrent effects against generalist herbivores. J Chem Ecol 21:507–523
- Vargas RR, Troncoso AJ, Tapia DH, Olivares-Donoso R, Niemeyer HM (2005) Behavioural differences during host selection between alate virginoparae of generalist and tobacco-specialist *Myzus persicae*. Entomol Exp Appl 116:43–53
- Wäschke N, Meiners T, Rostás M (2013) Foraging strategies of parasitoids in complex chemical environments. In: Wajnberg E, Colazza S (eds) Chemical ecology of insect parasitoids. Wiley, Chichester, UK
- Wäschke N, Hardge K, Hancock C, Hilker M, Obermaier E, Meiners T (2014) Habitats as complex odour environments: how does plant diversity affect herbivore and parasitoid orientation? PLoS One 9(1):e85152
- Zakir A, Sadek MM, Bengtsson M, Hansson BS, Witzgall P, Anderson P (2013) Herbivore-induced plant volatiles provide associational resistance against an ovipositing herbivore. J Ecol 101:410–417
- Zhang QH, Schlyter F (2003) Redundancy, synergism, and active inhibitory range of non-host volatiles in reducing pheromone attraction in European spruce bark beetle Ips typographus. Oikos 101:299–310